Protein labelling by 2-imino-1, 2-fluorenoquinone, a metabolite in vitro of the carcinogen N-(2-fluorenyl)acetamide

A mechanism for the interaction of the carcinogen N-(2-fluorenyl)acetamide with protein, which involves o-quinone imines, has recently been proposed. According to this mechanism, 2-imino-1,2-fluorenoquinone or 2-imino-2,3-fluorenoquinone, which are enzymic oxidation products in vitro of 2-amino-1-fluorenol or of 2-amino-3-fluorenol, respectively, combine with the appropriate functional group of the protein¹⁻³. This tentative conclusion was based on indirect spectrophotometric evidence². Recently, Cramer, Miller and Miller⁴ have discovered N-hydroxy-2-fluorenylacetamide as an important metabolite of N-(2-fluorenyl)acetamide⁴, and MILLER, MILLER AND HARTMANN consider that this metabolite may be the "proximate" carcinogen⁵. MILLER et al. have also advanced the view that a quinolimide, which may arise through a spontaneous rearrangement of N-hydroxy-2-fluorenylacetamide, may be the bound compound. We have therefore examined the labelling of bovine serum albumin by N-hydroxy-2- $[0^{-14}C]$ fluorenylacetamide as well as by the enzymic oxidation product of 2-amino-[1-14C]fluorenol-1. The results of this work indicate that only the latter is bound to the protein, while N-hydroxy-2-fluorenylacetamide, 2-aminofluorenol-1 and N-(1-hydroxy-2-fluorenyl)acetamide per se are not. The failure of N-hydroxy-2-fluorenylacetamide to combine with albumin also contradicts its spontaneous rearrangement to a quinolimide. Moreover, such a rearrangement was excluded by the absence of any phenolic compounds after Nhydroxy-2-fluorenylacetamide was incubated in acid or neutral media. Phenolic derivatives would be the end products of the spontaneous rearrangement of Nhydroxy-2-fluorenylacetamide to a quinolimide⁶. The data support the view, previously held^{2,3}, that the enzymic oxidation products of o-aminofluorenols play

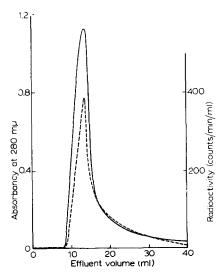


Fig. 1. Elution of the radioactivity and of the protein from a DEAE-cellulose column after incubating bovine serum albumin (0.15 μ mole) with 2-amino-[1-14C]fluorenol-1 (12.6 μ moles, 16900 counts/min/ μ mole) and cytochrome c - cytochrome oxidase. ———, absorbancy at 280 m μ ; ———, radioactivity.

a dominant role in the binding of the carcinogen N-(2-fluorenyl)acetamide to tissue proteins.

2-Amino-[1-14C]fluorenol-I (ref. 8) was oxidized by cytochrome c-cytochrome oxidase in the presence of bovine serum albumin. Alternatively, oxidation was accomplished by adding the aminofluorenol to a solution of K_3 Fe(CN)₆ and bovine serum albumin in phosphate buffer. The protein was then isolated by chromatography on N,N-diethyl aminoethyl cellulose columns, essentially as previously described³. Fig. I shows coincidence of radioactivity measurements and spectrophotometric protein determinations in the elution profile; this coincidence and the impossibility of extracting the radioactivity prove binding of the oxidized metabolite to albumin.

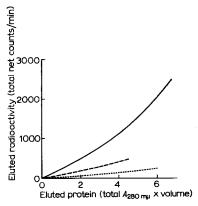


Fig. 2. Elution of the protein-bound radioactivity from DEAE-cellulose columns. The cumulative net radioactivity of the effluent fractions is plotted as the ordinate. The sum of the products of the absorbancy at 280 m μ and the volume of each fraction is plotted as the abscissa. ——, after incubating bovine serum albumin (0.15 μ mole) with cytochrome c – cytochrome oxidase and 2-amino-[1-14C]fluorenol-1 (12.6 μ moles, 16900 counts/min/ μ mole); ———, after incubating bovine serum albumin in the above system but omitting cytochrome c; ……, after incubating bovine serum albumin (0.15 μ mole) with N-hydroxy-2-[9-14C]fluorenylacetamide (1.87 μ mole, 16500 counts/min/ μ mole).

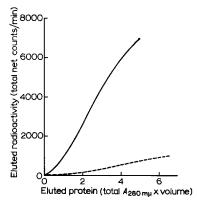


Fig. 3. Elution of the protein-bound radioactivity from DEAE-cellulose columns. The cumulative net radioactivity of the effluent fractions is plotted as the ordinate. The sum of the products of the absorbancy at 280 m μ and the volume of each fraction is plotted as the abscissa. ——, after the oxidation of z-amino-[1-MC]fluorenol-1 (11.8 μ moles, 21200 counts/min/ μ mole) by $K_3Fe(CN)_6$ in the presence of bovine serum albumin (0.15 μ mole); ——, after incubating bovine serum albumin in the above system but omitting $K_3Fe(CN)_6$.

N-hydroxy-2-[9-¹⁴C]fluorenylacetamide, prepared essentially by the published procedure⁴, was incubated in a phosphate buffer containing the bovine serum albumin and the protein was isolated as above. The results of this experiment and the binding of the o-quinoneimine are shown in Fig. 2. In separate experiments, N-hydroxy-2-fluorenylacetamide was incubated in buffers of pH I and 7.4. Ether extracts of these solutions were chromatographed⁴, and the chromatograms treated with p-dimethylaminobenzaldehyde and diazotized 7-nitro-2-fluorenamine⁴. The details of the foregoing experiments will be published at a later date.

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